

# Package: ICDS (via r-universe)

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**Type** Package

**Title** Identification of Cancer Dysfunctional Subpathway with Omics Data

**Version** 0.1.3

**Maintainer** Junwei Han <hanjunwei1981@163.com>

**Description** Identify Cancer Dysfunctional Sub-pathway by integrating gene expression, DNA methylation and copy number variation, and pathway topological information. 1)We firstly calculate the gene risk scores by integrating three kinds of data: DNA methylation, copy number variation, and gene expression. 2)Secondly, we perform a greedy search algorithm to identify the key dysfunctional sub-pathways within the pathways for which the discriminative scores were locally maximal. 3)Finally, the permutation test was used to calculate statistical significance level for these key dysfunctional sub-pathways.

**Depends** R (>= 4.3.0)

**Imports** igraph, graphite, metap, methods, org.Hs.eg.db

**Suggests** knitr, rmarkdown

**License** GPL (>= 2)

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.3.2

**VignetteBuilder** knitr

**Repository** <https://hanjunwei-lab.r-universe.dev>

**RemoteUrl** <https://github.com/hanjunwei-lab/icds>

**RemoteRef** HEAD

**RemoteSha** 599b89ae533fb207ad127826e0013805b2c6c2b0

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ICDS-package	<i>Identification of Cancer Dysfunctional Subpathway by integrating DNA methylation, copy number variation, and gene expression data</i>
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## Description

Identify Cancer Dysfunctional Subpathway by integrating gene expression, DNA methylation and copy number variation, and pathway topological information. 1)We firstly calculate the gene risk scores by integrating three kinds of data: DNA methylation, copy number variation, and gene expression. 2)Secondly, we perform a greedy search algorithm to identify the key dysfunctional subpathways within the pathways for which the discriminative scores were locally maximal. 3)Finally, the permutation test was used to calculate statistical significance level for these key dysfunctional subpathways.

## Author(s)

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Other contributors:

- Siyao Liu <liusiyao29@163.com> [contributor]

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combinep_three	<i>combinep_three</i>
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**Description**

'combinep\_three' combine three kinds of p-values,then,calculate z-score for them.

**Usage**

```
combinep_three(p1, p2, p3)
```

**Arguments**

p1	the p-values or corrected p-values
p2	the p-values or corrected p-values
p3	the p-values or corrected p-values

**Value**

A numeric vector of z\_scores

**Examples**

```
exp.p<-GetExampleData("exp.p")  
meth.p<-GetExampleData("meth.p")  
cnv.p<-GetExampleData("cnv.p")  
combinep_three(exp.p,meth.p,cnv.p)
```

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combinep_two	<i>combinep_two</i>
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**Description**

'combinep\_two' combine two kinds of p-values,then,calculate z-score for them.

**Usage**

```
combinep_two(p1, p2)
```

**Arguments**

p1	A numeric vector of p-values or corrected p-values
p2	A numeric vector of p-values or corrected p-values

**Value**

A numeric vector of z\_scores

**Examples**

```
exp.p<-GetExampleData("exp.p")  
meth.p<-GetExampleData("meth.p")  
combinep_two(exp.p,meth.p)
```

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coverp2zscore	<i>coverp2zscore</i>
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**Description**

'coverp2zscore' calculate z-scores for p-values

**Usage**

```
coverp2zscore(pdata)
```

**Arguments**

pdata            A numeric vector of p-values or corrected p-values

**Value**

A numeric vector of z\_scores

**Examples**

```
exp.p<-GetExampleData("exp.p")  
meth.p<-GetExampleData("meth.p")  
cnv.p<-GetExampleData("cnv.p")  
coverp2zscore(exp.p)  
coverp2zscore(meth.p)  
coverp2zscore(cnv.p)
```

---

envData	<i>The variables in the environment include an example expression profile,an methylation profile,an copy number variation data,amplified genes,deleted genes,A numeric vector of z_scores,p-values,A vector of 0/1s, indicating the class of samples,interested subpathways,Optimized subpathway,and the statistical significance p value and FDR for these optimal subpathways</i>
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### Description

Identify Cancer Dysfunctional Subpathway by integrating gene expression, DNA methylation and copy number variation, and pathway topological information. 1)We firstly calculate the gene risk scores by integrating three kinds of data: DNA methylation, copy number variation, and gene expression. 2)Secondly, we perform a greedy search algorithm to identify the key dysfunctional subpathways within the pathways for which the discriminative scores were locally maximal. 3)Finally, the permutation test was used to calculate statistical significance level for these key dysfunctional subpathways.

### Format

An environment variable

### Details

The environment variable includes the variable exp\_data, meth\_data, cnv\_data, amp\_gene, del\_gene, zzz, exp.p, meth.p, cnv.p

### Author(s)

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FindSubPath	<i>FindSubPath</i>
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### Description

'FindSubPath' uses a greedy search algorithm to search for key subpathways in each entire pathway.

### Usage

```
FindSubPath(
  zz,
  Pathway = "kegg",
  delta = 0.05,
  seed_p = 0.05,
  min.size = 5,
  out.F = FALSE,
  out.file = "Subpath.txt"
)
```

**Arguments**

zz	A numeric vector of z_scores.
Pathway	The name of the pathway database.
delta	Diffusion coefficient in each step of searching subpath.
seed_p	Define gene whose p-value smaller than seed_p as seed gene.
min.size	The smallest size of subpathways.
out.F	Logical,tell if output subpathways.
out.file	file name of subpathways.

**Value**

Key dysfunctional subpathways in each pathway, in which the risk score of the genes were significantly higher.

**Examples**

```
require(graphite)
zz<-GetExampleData("zzz")
k<-FindSubPath(zz)
```

---

getCnvp

*getCnvp*


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**Description**

‘getCnvp’ perform t-test on copy number variation data

**Usage**

```
getCnvp(
  exp_data,
  cnv_data,
  amp_gene,
  del_gene,
  p.adjust = TRUE,
  method = "fdr"
)
```

**Arguments**

exp_data	A data frame
cnv_data	Copy number variation data
amp_gene	A vector of strings, the IDs of amplified genes.
del_gene	A vector of strings, the IDs of deleted genes.
p.adjust	Logical,tell if returns corrected p-values
method	Correction method,which can be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY",

**Details**

cnv\_data is TCGA level4 data.if p.adjust=TRUE,return corrected p-values,if p.adjust=FALSE,return p-values

**Value**

A numeric vector of p-values or corrected p-values

**Examples**

```
exp_data<-GetExampleData("exp_data")
meth_data<-GetExampleData("meth_data")
cnv_data<-GetExampleData("cnv_data")
amp_gene<-GetExampleData("amp_gene")
del_gene<-GetExampleData(("del_gene"))
getCnvP(exp_data, cnv_data, amp_gene, del_gene, p.adjust=FALSE, method="fdr")
```

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GetExampleData	<i>Get the example data</i>
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**Description**

Get the example data of test package for litte trials.

**Usage**

```
GetExampleData(exampleData)
```

**Arguments**

exampleData      A character, should be one of "exp\_data", "meth\_data", "cnv\_data", "amp\_gene", "del\_gene", "label1", "label2", "zz", "exp.p", "meth.p", "cnv.p" and "pathdata".

**Details**

The function `getExampleData(ExampleData = "exp.p")` obtains a vector of lncRNAs confirmed to be related with breast cancer. The function `getExampleData(ExampleData = "Profile")` obtains the expression pr

**References**

Subramanian, A., Tamayo, P., Mootha, V.K., Mukherjee, S., Ebert, B.L., Gillette, M.A., Paulovich, A., Pomeroy, S.L., Golub, T.R., Lander, E.S. et al. (2005) Gene set enrichment analysis: a knowledgebased approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A*, 102, 15545-15550.

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`getExpp`*getExpp*

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**Description**

'getExpp' perform t-test on Expression profile data

**Usage**

```
getExpp(exp_data, label, p.adjust = TRUE, method = "fdr")
```

**Arguments**

<code>exp_data</code>	A data frame, the expression profile to calculate p-value for each gene, the row-names should be the symbol of genes.
<code>label</code>	A vector of 0/1s, indicating the class of samples in the expression profile, 0 represents case, 1 represents control.
<code>p.adjust</code>	Logical, tell if returns corrected p-values
<code>method</code>	Correction method, which can be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY",

**Details**

For a given expression profile of two conditions, ICDS package provide t-test method to calculate p-values or corrected p-values (if `p.adjust=TRUE`, return corrected p-values, if `p.adjust=FALSE`, return p-values.) for each genes. The row of the expression profile should be gene symbols and the column of the expression profile should be names of samples. Samples should be under two conditions and the label should be given as 0 and 1.

**Value**

A numeric vector of p-values or corrected p-values

**Examples**

```
profile<-GetExampleData("exp_data")
label<-GetExampleData("label1")
getExpp(profile, label, p.adjust=FALSE)
```



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getMethp	<i>getMethp</i>
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### Description

'getMethp' perform t-test on Methylation profile data

### Usage

```
getMethp(meth_data, label, p.adjust = TRUE, method = "fdr")
```

### Arguments

meth_data	A data frame, the Methylation profile to calculate p-value for each gene, the rownames should be the symbol of genes.
label	label A vector of 0/1s, indicating the class of samples in the Methylation profile, 0 represents case, 1 represents control.
p.adjust	Logical,tell if returns corrected p-values
method	Correction method,which can be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY",

### Details

For a given Methylation profile of two conditions, ICDS package provide t-test method to calculate p-values or corrected p-values(if p.adjust=TRUE,return corrected p-values,if p.adjust=FALSE,return p-values.) for each genes. The row of the Methylation profile should be gene symbols and the column of the Methylation profile should be names of samples. Samples should be under two conditions and the label should be given as 0 and 1.

### Value

A numeric vector of p-values or corrected p-values

### Examples

```
profile<-GetExampleData("meth_data")
label<-GetExampleData("label2")
getMethp(profile,label,p.adjust=FALSE)
```

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opt_subpath	<i>opt_subpath</i>
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### Description

‘opt\_subpath’ Optimize interested subpathways.If the number of genes shared by the two pathways accounted for more than the Overlap ratio of each pathway genes,then combine two pathways.

### Usage

```
opt_subpath(subpathdata, zz, overlap = 0.6)
```

### Arguments

subpathdata	interested subpathways
zz	a vector of z-scores
overlap	Overlap ratio of each two pathway genes

### Value

Optimized subpathway:the number of genes shared by any two pathways accounted for less than the Overlap ratio of each pathway genes.

### Examples

```
zz<-GetExampleData("zzz")
subpathdata<-GetExampleData("subpathdata")
optsubpath<-opt_subpath(subpathdata,zz,overlap=0.6)
```

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Permutation	<i>Permutation</i>
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### Description

the permutation test method 1 and method 2 were used to calculate the statistical significance level for these optimal subpathways.

### Usage

```
Permutation(
  subpathwayz,
  zz,
  nperm1 = 1000,
  method1 = TRUE,
  nperm2 = 1000,
  method2 = FALSE
)
```

**Arguments**

subpathwayz	Optimize interested subpathways
zz	a vector of z-scores
nperm1	times of permutation to perform use method1
method1	permutation analysis method1
nperm2	times of permutation to perform use method2
method2	permutation analysis method2

**Value**

the statistical significance p value and FDR for these optimal subpathways

**Examples**

```
require(graphite)
keysubpathways<-GetExampleData("keysubpathways")
zzz<-GetExampleData("zzz")
Permutation(keysubpathways, zzz, nperm1=10, method1=TRUE, nperm2=10, method2=FALSE)
```

---

PlotSubpathway

*PlotSubpathway*


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**Description**

PlotSubpathway:plot a network graph when user input a list of gene

**Usage**

```
PlotSubpathway(
  subpID,
  pathway.name,
  zz,
  Pathway = "kegg",
  layout = layout.fruchterman.reingold
)
```

**Arguments**

subpID	gene list of a interested subpathway
pathway.name	name of the interested subpathway
zz	z-score of each gene
Pathway	the name of the pathway database
layout	The layout specification( <a href="#">layout_</a> ). It must be a call to a layout specification function.

**Value**

Network graph

**Examples**

```
require(graphite)
```

```
subpID<-unlist(strsplit("ACSS1/ALDH3B2/ADH1B/ADH1A/ALDH2/DLAT/ACSS2","/"))  
pathway.name="Glycolysis / Gluconeogenesis"  
zzz<- GetExampleData("zzz")  
PlotSubpathway(subpID=subpID,pathway.name=pathway.name,zz=zzz)
```

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